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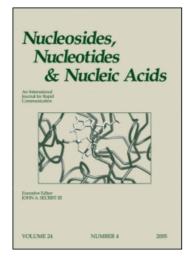
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Nucleosides, Nucleotides and Nucleic Acids

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Synthesis, Solution Conformation and Biological properties of 2',3'-Dideoxy-3'-fluoro-D-*erythro*-pentofuranosides of 2-Thiouracil and 2-Thiothymine

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SYNTHESIS, SOLUTION CONFORMATION AND BIOLOGICAL PROPERTIES OF 2',3'-DIDEOXY-3'-FLUORO-D-ERYTHRO-PENTOFURANOSIDES OF 2-THIOURACIL AND 2-THIOTHYMINE

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Abstract. The synthesis of the α - and β -anomers of 2',3'-dideoxy-3'-fluoro-2-thiouridine and 2',3'-dideoxy-3'-fluoro-2-thiothymidine *via* Lewis acid catalysed nucleoside condensation is described. High resolution ¹H NMR data, solution conformations and biological properties are also presented.

2',3'-Dideoxy-3'-fluorothymidine (FLT) is one of the more potent *in vitro* inhibitors of HIV and its reverse transcriptase, but exhibits marked cytotoxicity. Its 4-thio derivative exhibits an enhanced therapeutic index in MT-4 cells, ¹ prompting us to prepare the corresponding 2-thio analogues 2',3'-dideoxy-3'-fluoro-2-thiouridine (5b) and 2',3'-dideoxy-3'-fluoro-2-thiothymidine (7b) and their α -anomers 5a and 7a.

Condensation of silylated 2-thiouracil (1) with 1,5-di-O-benzoyl-2,3-dideoxy-3-fluoro- β -D-erythro-pentofuranose (3b), synthesized as previously described, in the presence of SnCl₄ (molar ratio of reagents 2:1:3) was conducted in 1,2-dichloroethane-acetonitrile (1:1, v/v) for 2 h at 30°C, followed by standard work-up. From a mixture of the reaction products in methanol, the α -anomer 4a was deposited as crystals. Column chromatography of the remaining mixture yielded an additional quantity of 4a (total, 47%), and the β -anomer 4b (16%). The overall yield of products was improved when the amount of SnCl₄ was increased to 4 molar equivalents, but this lead to a decrease in the ratio β/α . The same trend was observed on prolongation of the reaction time. Deblocking of 4a and 4b with methanolic ammonia yielded the free nucleosides 5a and 5b, respectively.

Quite unexpectedly, replacement of silylated 2-thiouracil by silylated 2-thiothymine in the condensation reaction led to a complex mixture of products. Reaction of 2 with 3b in the presence of trimethylsilyl triflate (TMS-Tfl), with a molar ratio of reactants of 2:1:1.5, in

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TABLE 1

1 H NMR (500 Mhz) data of 2',3'-dideoxy-3'-fluoro-2-thiouridines 5a, b and 7a, b and their analogues

			_							
Compound	Chemical shifts (ppm) in D ₂ O vs internal DSS									
	1'	2'	2"	3'	4'	5'	5"	5	6	
ß-3'F dT	6.35	2.39	2.65	5.36	4.38	3.38	3.82		7.68	
ß-3'F dU	6.42	2.46	2.77	5.42	4.48	3.88	3.88		7.94	
5a	6.74	2.81	2.55	5.25	4.80	3.63	3.66	5.96	7.77	
7a	6.67	2.73	2.43	5.17	4.80	3.59	3.55		7.50	
α -2 S dU	6.78	2.90	2.41	4.51	4.64	3.80	3.72	6.78	8.09	
5b	7.01	2.11	2.81	5.27	4.33	3.80	3.80	5.97	8.16	
7b	6.95	2.03	2.67	5.19	4.23	3.73	3.73		7.97	
ß-2 S dU	7.03	2.38	2.70	4.52	4.19	3.97	3.88	7.03	8.12	
Compound	Proton-proton coupling constant (Hz)									
	1',2'	1',2"	2',2"	2',3'	2",3	' 3	',4'	4',5'	4',5"	
β-3'F dT	9.29	5.81	-14.84	5.31	0.60	0	.87	4.43	4.16	
ß-3'F dU	9.13	5.86	-14.97	5.01	0.55	0	.75	4.37	4.37	
5a	7.09	1.22	-15.87	4.71	0.80	0	.80	4.57	3.62	
7a	7.13	1.14	-15.86	4.78	1.00	1	.00	4.36	3.63	
α-2 S dU	6.90	1.80	-15.44	5.93	1.80	1	.80	4.02	5.42	
5b	8.73	5.51	-14.60	5.00	1.00	1	.00	3.14	3.14	
7b	8.73	5.54	-14.64	5.15	1.00	1	.00.	2.78	2.78	
ß-2 S dU	6.59	6.28	-14.25	6.62	4.43	4	.05	3.37	5.00	

acetonitrile for 4 h at room temperature, resulted in only slow formation of nucleoside products (monitored by TLC). Addition to this mixture of a further 1.5 molar equivalent of TMS-Tfl, and continuation of the reaction for 2 h, led to isolation, after standard work-up and column chromatography, of a non-separable mixture of **6a**, **b** (45% relative to **3b**, β/α 1:3 by 1 H NMR), the α -anomer **3a** of the starting sugar (11%), identified with an authentic sample, unaltered **3b** (20%), and other unidentified products. Subsequent condensation of **2** with **3b** in the presence of TMS-Tfl (molar ratio 2:1:3) in acetonitrile for 24 h at room temperature afforded a mixture of **6a**, **b** (23%, β/α 1:3), and an enhanced proportion of unidentified products, but not including **3a**, **b** (TLC). Replacement of TMS-Tfl by SnCl₄ did not lead to an improvement in either the yield of **6a**, **b** (40%) or the β/α ratio (1:4 by 1 H NMR). These and the blocked anomers **6a**, **b** could not be separated. However, standard deprotection of the mixture of blocked anomers, followed by column chromatography, afforded the desired free

Compound	S	g+	t	g-	Ng+	Nt	Sg+	St	Sg-
ß-3'F dT	1.02	0.51	0.24	0.24			0.53	0.25	0.24
ß-3'F dU	1.04	0.50							
5a	1.03	0.56	0.18	0.26			0.58	0.19	0.26
7a	1.00	0.58	0.19	0.23			0.58	0.19	0.23
α -2 S dU	0.90	0.42	0.40	0.19	0.06	0.04	0.35	0.36	0.19
5b	1.00	0.75							
7b	1.00	0.81							
ß-2 S dU	0.60	0.52	0.37	0.11	0.25	0.15	0.27	0.22	0.11

TABLE 2
Conformer populations in D₂O

nucleosides 7a and 7b, whose structure was confirmed by high resolution ¹H NMR (Table 1), CD and MS spectroscopy.

Their conformational properties in aqueous medium are of some interest (Table 2). Both anomers of 5 and 7 exhibit a virtually 100% population of the S conformer of the pentose moiety. By contrast, in the absence of the 3'-fluoro-, the conformation of the pentose ring is dependent on the anomeric form, viz. about 60% S for 2-thio-2'-deoxyuridine and 90% for its α -anomer, and 60% S for 2-thiothymidine and 80% S for its α -anomer.

Biological Results

Nucleosides **5b** and **7b** were evaluated against HIV-1(III_B) and HIV-2(ROD) strains in CEM cells, and were found to be inhibitory at an EC₅₀ of 3 μ g/ml for **5b**, and 0.07 to 0.10 μ g/ml for **7b**. CC₅₀in CEM cells was 100 μ g/ml for **5b** and 58 μ g/ml for **7b**.

Nucleosides **5b** and **7b** were shown to be phosphorolytically cleaved by thymidine phosphorylase (dThd Pase) from *E.coli* in contrast to FLT which was not a substrate for dThd Pase at the conditions used (4 mM substrate concentration, 50 mM phosphate buffer pH 7.4 and dThd-Pase, $1.66 \text{ U}/100 \mu\text{l}$).

Catabolism of **5b** and **7b** as well as the lack of thymidine kinase and/or of reverse transcriptase affinity may be the reason for the lack of activity of these compounds. Further biological investigations are underway.

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References

- 1. Matthes, E.; Lehman, C.; von Janta-Lipinski, M.; Scholz, D. Biochem. Biophys. Res. Commun., 1989, 165, 488.
- Mikhailopulo, I.A.; Pricota, T.I.; Poopeiko, N.E.; Klenitskaya, T.V.; Khripach, N.B., Synthesis, 1993, 700.